BIOSYNTHESIS OF IODININ: INCORPORATION OF D-[1-
$$^{14}$$
c]-, D-[6- $^{14}$ c]-
AND D-[1,6,7- $^{14}$ c<sub>2</sub>]-SHIKIMIC ACID

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Earlier results from this laboratory agve the distribution of D-[6-14C]shikimic acid (as 1) label in iodinin (2) as 22% in C-1, C-4, C-6, and C-9 and 74% in C-4a, C-5a, C-9a, and C-10a. These results were at variance with those reported for iodinin (12% and 104%) and phenazine-1-carboxylic acid (3) (1% and 102%) from the same precursor; both sets were incompatible with earlier results on iodinin derived from DI-[1,6-14C2]shikimic acid (50% and 25%, the remaining 25% being in C-2, C-3, C-7, and C-8).

Our distribution was interpreted as the label in ring A being confined to C-10a, and that in ring B being equally divided over C-5a and C-9 (or C-6 and C-9a) which would give a distribution of 25 and 75%. To resolve the two possible modes of symmetrisation of ring B (shikimic acid C-6 becoming equivalent to C-2 or C-4) we have now fed D- $\left[1-\frac{14}{3}\right]$  shikimic

acid; results are given in the table. Using our previous degradation route (2)—(4)
—(5)—(6) and methods, up to 48% of the activity of (5) was unaccounted for in the pyrazine and BaCO<sub>3</sub>. The explanation of this apparent loss probably lies in the copper chromite reagent used in the last decarboxylative step (5)—(6) causing degradation of some of the pyrazine ring to fragments which were collected and counted as BaCO<sub>3</sub>, in this case active BaCO<sub>3</sub> being diluted with inactive material. Using copper-bipyrid-yl-quinoline<sup>5</sup> (CuQ) as the decarboxylation reagent 100% of this label was found in the carbon dioxide of step (5)—(6) corresponding to C-1, C-4, C-6 and C-9 of iodinin (2).

Degradation of the pyrazine ring, now labelled, by the copper chromite reagent also accounts for the appreciable activity found in C-1, C-4, C-6 and C-9 of iodinin using D-[6-<sup>14</sup>C]shikimic acid<sup>1,2</sup> (although the same degradation applied to phenazine-1-carboxylic acid had given only 1% of the label in these positions<sup>2</sup>). Repetition of our earlier work<sup>1</sup> but with CuQ decarboxylation now gave consistent results with only a small activity in the BaCO<sub>3</sub>; careful drying of the hygroscopic N,N-diacetylpiperazine used for the determination of the activity of the pyrazine gave 100% of the activity in C-4a, C-5a, C-9a and C-10a of the phenazine system. These results, together with the label distribution in iodinin

derived from D-[1,6,7-14C<sub>3</sub>]shikimic acid<sup>4</sup> (see table) are not only self consistent but also substantiate Hollstein and McCamey's conclusion that shikimic acid (numbering) relates to the phenazine nucleus as in (7),<sup>2</sup> in iodinin as well as in phenazine-1-carboxylic acid<sup>2</sup> and pyocyanin.<sup>2,6</sup>

TABLE

Distribution of activity as % of phenazine-1,6-diol

14C-Labelled Shikimate	Incorporationa	Pyrazinetetracarboxylic	Pyrazine	Carbon Dioxide <sup>C</sup>
precursor	%	acid b		
1	42	100	o	52, 67 <sup>d</sup>
		102	0	100, 101 <sup>e</sup>
6	31	102	113 <sup>f</sup> 100 <sup>g</sup>	1.2 <sup>e</sup>
1,6,7	34 <sup>h</sup>	99	55 <sup>f</sup> 47 <sup>g</sup>	49 <sup>e</sup>
			56 <sup>f</sup> 50 <sup>g</sup>	50 <sup>e</sup>

- a. Based on phenazine-1,6-diol.
- b. Radioactivity determined on tetramethyl ester.
- c. Radioactivity determined on BaCO<sub>3</sub> in scintillator.
- d. Decarboxylation by heating with copper chromite.
- e. Decarboxylation by heating with Cu, bipyridyl and quinoline.
- f. Radioactivity measured on pyrazine assayed by u.v.
- g. Radioactivity determined on N, N-diacetylpiperazine.
- h. Allowing for loss of  $\frac{1}{5}$  of the activity.

## REFERENCES

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- 5. Cepper powder (20 mg), 2,2'-bipyridyl (5 mg), and pyrazinetetracarboxylic acid (14 mg) in CO<sub>2</sub>-free quinoline (1.5 ml) heated in a bath at 210° for 2 hr in a stream of CO<sub>2</sub>-free nitrogen, which then passed through, successively, saturated aqueous mercuric chloride (2 traps) and aqueous barium hydroxide (38 g/l; 2 traps).
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